

REVIEW

Open Access



Topical biomaterials to prevent post-tonsillectomy hemorrhage

Lumei Liu¹, Cole Rodman², Noah E. Worobetz^{1,3}, Jed Johnson⁴, Charles Elmaraghy^{2,3} and Tendy Chiang^{1,2,3*}

Abstract

Despite advances in surgical technique, postoperative hemorrhage remains a common cause of mortality and morbidity for patients following tonsillectomy. Application of biomaterials at the time of tonsillectomy can potentially accelerate mucosal wound healing and eliminate the risk of post-tonsillectomy hemorrhage (PTH). To understand the current state and identify possible routes for the development of the ideal biomaterials to prevent PTH, topical biomaterials for eliminating the risk of PTH were reviewed. Alternative topical biomaterials that hold the potential to reduce the risk of PTH were also summarized.

Keywords: Post-tonsillectomy hemorrhage, Biomaterial, Topical, Surgical hemostasis, Tonsillectomy

Background

Tonsillectomy is among the most common surgeries performed in pediatric patients. Tonsillectomy is primarily indicated for children who suffer from recurrent infection of the tonsils and / or for tonsillar enlargement that contributes to airway obstruction and sleep disturbance. Over 530,000 children under the age of 15 undergo tonsillectomy annually in the United States [1–3]. Following surgical removal of the tonsils, the surgical site(s) (tonsillar fossae) heal secondarily. One of the most common complications following tonsillectomy is hemorrhage from the tonsillar fossae, called post-tonsillectomy hemorrhage (PTH) [4]. This can either occur within the first 24 h (primary PTH) or from > 24 h to 2 weeks after surgery (secondary PTH). PTH rates (2.5–7%) have been increasing according to recent statistics: there are an estimated 25,000 cases of PTH in the United States each year, while in 1995 that number was 4300 [5]. Post-tonsillectomy hemorrhage often requires hospital readmission, surgical intervention, and can even result in significant morbidity such as shock, airway obstruction and need for blood transfusion [6]. The hemorrhage is generally unexpected and unpredictable [7–12], and resuscitation with life-threatening PTH can prove especially challenging [13, 14].

Attempts to reduce the morbidity of tonsillectomy have included changes to the surgical approach and peri-operative medications to reduce the risk of bleeding. Technical modifications to the procedure, such as partial or intracapsular tonsillectomy, can reduce pain and PTH [15–17]. However, such modifications are not indicated for chronic infection and can result in tissue regrowth, requiring revision surgery [18, 19].

A low cost, easily applied, effective strategy to prevent PTH remains elusive. Topical biomaterials have many appealing properties, making them a potentially ideal strategy for PTH prevention. Biomaterials can be fabricated with adhesive properties, and once adherent, they provide physical barriers to protect healing tissue from complex shear forces created by cough and swallowing. Some biomaterials can confer antimicrobial properties which facilitate the PTH prevention by protecting tonsil wounds from infection. An ideal topical biomaterial can prevent PTH by accelerating wound healing and remucosalization, and by expediting and amplifying the coagulation cascade and clot formation. Additionally, a topical therapy can include analgesic substances to locally reduce pain.

The proposed mechanism of secondary hemorrhage is retraction and sloughing of the eschar covering the healing tonsillar bed [20]. The tonsillar bed is at especially high risk for secondary hemorrhage due to the complexity of the tonsil environment, the lack of self-compression by surrounding tissues, and the robust

* Correspondence: Tendy.Chiang@NationwideChildrens.org

¹Center of Regenerative Medicine, Research Institute at Nationwide Children's Hospital, Columbus, OH, USA

²College of Medicine, The Ohio State University, Columbus, OH, USA
Full list of author information is available at the end of the article



vascular supply to the area. Thus, wound healing and clot stabilization is a critical process in preventing PTH. Biomaterials preventing PTH through improving wound healing after tonsillectomy have been poorly reported. Many studies evaluated surgical outcomes of biomaterials including pain, hemorrhage severity, and incidence of bleeding, but there are few standard methods to evaluate their impact on wound healing after tonsillectomy [21]. An ideal, functional biomaterial is expected to 1) facilitate hemostasis intraoperatively by improving coagulation and clot formation; 2) stabilize the clot and provide protection or compression within the first 24 h, thus preventing primary PTH; 3) improve wound healing and remucosalization, shortening the at-risk time from secondary PTH; 4) provide a physical barrier for the duration of the wound healing process.

Since the mid-90s', numerous studies have been devoted to the study of topical biomaterials (sulfathiazole gum) to prevent PTH [22]. We present a comprehensive review of these topical biomaterials, categorizing them into fibrin-based, gelatin-based, cellulose-based, bismuth subgallate-based, antifibrinolytics, and natural materials. We also include other promising biomaterials as potential target therapies for PTH prevention. These biomaterials have specific properties to prevent PTH to some degree, however, some limitations and side effects impeded their application in reality, for example high cost (fibrin-based patch), inconsistent clinical outcomes (fibrin-based sealant and gelatin-based matrix), risk of suffocation (cellulose-based hemostats), non-reduced pain (cellulose-based hemostats secured with suture) etc. We hope to illuminate the current state of topical methods of PTH prevention and offer a course of future directions to develop biomaterial for PTH prevention.

Main text: topical biomaterials used for PTH

Search strategy

All topical biomaterials used for PTH were searched in databases of Google Scholar and the Food and Drug Administration (FDA). The key words used to identify related topics include: post-tonsillectomy hemorrhage, biomaterial, topical, hemostasis, tonsillectomy. Articles and patents published and searchable in google scholar after the year of 2000 were reviewed. The FDA approval status of identified biomaterials for PTH were found in FDA website. The reference lists of all relevant primary studies, review articles and patents.

Fibrin-based biomaterials

Fibrin is a common biomaterial that has been widely applied as a hemostatic sealant in clinical and bio-engineering fields [23–26]. The hemostasis and wound healing mechanism of fibrin-based biomaterial follows

the principles of physiological fibrin clot formation. Most fibrin-based biomaterials contain fibrinogen and thrombin (Quixil® does not contain fibrinogen). Thrombin (a clotting enzyme) transforms fibrinogen to fibrin monomers when combined with calcium and factor XIII from wound surfaces. This initiates polymerization of fibrin monomers to polymeric fibrin clots. Fibrin clots are large mesh-like polymers that trap platelets and red blood cells, which in turn release more clotting factors amplifying the cycle. When procedures for wound closure are limited, clots are relied upon to achieve tissue sealing and hemostasis in several surgical procedures [27]. Thrombin (Factor IIa) initiates cleavage of fibrinogen into fibrin monomers, accelerating clot formation [28, 29]. Meanwhile thrombin activates factor XIII, which stabilizes coagulated fibrin by covalently crosslinking [30]. Thrombin also acts directly on platelets, factors V, VII, and X, XII, and smooth muscle cells, leading to hemostasis and wound healing [31, 32].

Current forms of fibrin-based sealant include liquid fibrin-based glue and a fleece/patch. The first fibrin-based glue product approved by Food and Drug Administration (FDA) was Tisseel® fibrin sealant glue (Baxter International, Deerfield, IL) [33]. Later, fibrin glue products such as Tissucol® (Baxter International) and Quixil® (OMRIX biopharmaceuticals, Inc., New York City, NY) were used in tonsillectomy patients [34, 35]. Application of these fibrin glues had no effect on post-tonsillectomy pain control and bleeding prevention [34, 35]. In a review of hemostatic glues in tonsillectomy, it was concluded that fibrin glues were not recommended for routine use in current clinical practice as they were not effective in reducing severity of pain and bleeding [31]. However, other studies showed beneficial outcomes of fibrin-based glue for PTH prevention. Quixil tested in a prospective randomized study reduced pain and prevented bleeding in tonsillectomy patients compared with bipolar or needle point electrocautery (0% vs. 4.35%) [36]. Another prospective randomized single-blind study proved Quixil can decrease immediate inflammatory response in patients after tonsillectomy [36, 37]. In a prospective randomized study of 40 patients, another liquid form of fibrin sealant (spray) reduced pain compared with electrocautery hemostasis alone [38]. Evicel® (Ethicon Inc., Somerville, NJ) is a new formulation of previously available fibrin sealant Quixil (EU) or Crosseal™ (US), and it has been used as an effective hemostat in orthopedic [39], endonasal surgeries [40, 41], and epistaxis management [42, 43] but benefit in tonsillectomy has yet to be assessed.

Compared with fibrin glue, fibrin-based patch or fleece has demonstrated advantages to be used for PTH prevention. Compared to the complex manipulation of fibrin glue, which requires storage at low temperatures

and long thawing time [31], fibrin-based patches are ready-to-use. They do not require preparation, mixing, moistening with saline or refrigeration. Another reason fibrin-based glues are not ideal PTH prevention is that they lack the necessary mechanical strength to perform as an adhesive or tissue sealant [44]. A patch/fleece provides a robust and solid mechanical support to fibrin glue, making the fibrin-based patch an effective sealant. For example, one fibrin-based fleece product, TachoComb® (Nycomed Pharma, Switzerland) is comprised of a collagen sponge and a dried layer of fibrinogen and thrombin. TachoComb is a widely used method that has been used in diffuse bleeding from parenchymatous organs or bleeding of the lung both in conventional and endoscopic surgery [45]. One prospective study was performed to assess TachoComb in children for postoperative complications after tonsillectomy ($n = 53$ for treatment group and $n = 57$ for control group). It was demonstrated the fibrin-based patch significantly reduced pain and post-surgery bleeding from 8.8 to 0%. [46]. While in another prospective study, the rate of postoperative hemorrhage and re-admission for PTH in TachoComb treatment group were not significant different with control group [47].

Compared with TachoComb, TachoSil® (Takeda Pharmaceutical Company, Tokyo, Japan) is an FDA approved biomaterial that is non-aprotinin and contains human thrombin instead of the bovine thrombin found in TachoComb [47–49]. Common ingredients in both TachoSil and TachoComb patches is human fibrinogen (5.5 mg/cm^2), and additional active ingredients in TachoComb is bovine aprotinin [50], which could explain different outcome in future comparative studies. TachoSil is recommended for use in small wounds due to the poor sealant properties in large wounds [51]. TachoSil has been used in a variety of abdominal surgeries including hepatic, pancreatic, spleen, gastrointestinal, and inguinal hernia repair [52]. However, it was shown that TachoSil has a risk of adhesive failure compared with another fibrin sealant patch, Evarrest® (Ethicon), in a swine spleen incision model [53]. Evarrest has been approved by FDA [54], and has been used to treat severe soft tissue surgical bleeding such in abdominal, pelvic, and retroperitoneal sites, and in non-cardiac thoracic, liver, cardiovascular surgeries [55–57]. Although Evarrest has not been tested with tonsillectomies, its application in these surgeries and relevant results would encourage the use of Evarrest for PTH.

Some fibrin-based biomaterial products contain tranexamic acid (TXA) instead of aprotinin as an antifibrinolytic adjuvant. Of note, TXA is potentially neurotoxic [41]. In 2010, TachoComb was evaluated in pigs that the aprotinin component caused anaphylactic relations and risk of renal failure [47]. It was also too expensive for

some surgical units [58]. The high cost of fibrin-based patches might limit the clinical application of this biomaterial [59, 60]. Despite improvement in PTH rates demonstrated in Quixil, TachoComb, TachoSil, and Evarrest, application has not been widely adopted due to 1) Lack of persistence for entire bleeding risk interval; 2) Anaphylaxis; 3) Attachment failure; 4) Not tested yet; 5) High cost.

Gelatin-based biomaterials

Gelatin-thrombin hemostatic matrix (GTHM) agent is a gelatin-based biomaterial consisting of bovine-derived gelatin matrices and human-derived thrombin. It has been used to treat and manage secondary PTH [61]. Gelatin-based materials provide a combination of two independent hemostatic agents, gelatin and thrombin, in order to achieve hemostasis [62, 63]. Gelatin matrix granules swell to produce a tamponade effect for wound healing [28, 64]. Upon delivery to the wound site, it serves as a substrate for platelet adhesion and fibrin activation, leading to clot formation. Thrombin accelerates and stabilizes fibrin clot formation, improving hemostasis and wound healing [31, 32].

The absorbable liquid form product (Table 1), FloSeal™ Hemostatic Matrix (Baxter International) was approved by the FDA [72]. FloSeal has been advocated to improve intraoperative hemostasis in cardiac and spinal surgeries [63, 90], transphenoidal pituitary surgery [91], the management of epistaxis [92], endoscopic sinus surgery [93], and is commonly used in otolaryngologic interventions by otolaryngologists [94]. FloSeal was tested in a randomized controlled trial on 68 pediatric patients undergoing adenotonsillectomy. Results showed that FloSeal was safe and efficacious, and decreased postoperative morbidity and blood loss comparing with electrocautery hemostasis after cold steel adenotonsillectomy [67]. In a later clinical trial, FloSeal showed reduction in pain medication, promoted mucosal recovery and faster wound healing (less thickness of wound plaques), but no significant reduction in postoperative hemorrhage [69]. Another study concluded that FloSeal on ligatured fossa had no effect on pain reduction comparing with control group (ligatured fossa without FloSeal) [68]. Absorbable gelatin medicated sponges or powders [95, 96] (Table 2) are another type of gelatin-based hemostat, which have not been used for PTH yet. They are discussed in section two.

The clinical trials of gelatin-based biomaterials for PTH have inconsistent outcomes on pain, hemorrhage severity and incidence. The other side effects of gelatin-based hemostatic agents include 1) A nidus for infection and abscess formation, as they have been reported to potentiate bacterial growth, 2) foreign body reactions and “encapsulation” of fluid, and 3) toxic shock syndrome,

Table 1 Summary of topical biomaterials to prevent PTH

Biomaterial	Decrease PTH severity (Y/N)	Decrease PTH incidence (Y/N)	Reduce pain (Y/N)	Study type	Commercial Products	Administer type	Other outcomes	FDA approval for human (Y/N)
Fibrin-based Hemostasis glue/fleece/patch	N [31, 34, 35]	Y [36]	N [31, 34, 35], Y (spray) [38]	randomized double-blind study on 50 adults patients [34], prospective randomized double-blind study on 168 consecutive patients and systematic review [31]	Tissuol™/Tisseel™, and Crosseal™ / Quixil®, Evicel®	Post-op	Easy to use, maybe time efficient in operation but storage at low temperatures, and long thawing time [31]. decreased immediate inflammatory response following adenotonsillectomy [37]. Not enough sample sizes were studied [31].	Y [59, 65]
Fibrin-based collagen fleece/patch	reduced emergency surgery for severe PTH without an apparent adverse effect [66]	N [66]	Y [66]	Clinical study on 1057 children patients [66]	TachoComb®, TachoSil®, Evarrest® fibrin sealant patch	Post-op	provided a mechanical scaffold on which vascular regeneration occurred. Not cost efficient. Too expensive for some surgical units [58]	Y TachoSil [48] Y Evarrest® [54] N TachoComb
Gelatin-based Hemostat	Y [67] N	Y [61], [67]	N [68]	Case study by reviewing 42 pediatric patients and retrospective data analysis [61], randomized, controlled trial in adults [68, 69] and children [67, 70] and a review [71]	FloSeal™ Hemostatic Matrix	Post-op [61, 71]	simple, safe, and efficacious and cost-effective [61, 70]	Y [72]
Absorbable cellulose-based Hemostat	Y [73]	Y (when combine with saturation) [74]	N (when combine with saturation) [74]	Surgical plus saturation on total of 760 patients (393 males, 367 females) between the ages of 4 and 35 years [74]	Surgicel®, Cellistyp™, Puhacel®, Oxycel®, Gelita®, GuraTamp®	Intra-op	aids in clot formation by activation of the extrinsic and intrinsic coagulation pathways. Introduced a risk of aspiration and suffocation [75]; Possible inflammation [76].	Y Surgicel® [77] N for others
Bismuth subgallate-based	Y [78]	Y (BSG mixed with phenylephrine hydrochloride) [79]	N [81]	Clinical study on patients underwent tonsillectomy [78, 81]	Spectrum Chemical (powder), mixing 26 g of BSG powder to 20 ml of normal saline with 0.7 ml of 1:1000 adrenaline	Post-op	decreases operating time by significantly reducing the hemostasis time and the number of ligatures [78, 81]; accelerates the cascade of blood clotting [79]; accelerate the intrinsic clotting pathway through the activation of factor XII (Hageman factor) [82].	Y [62]

Table 1 Summary of topical biomaterials to prevent PTH (*Continued*)

Biomaterial	Decrease PTH severity (Y/N)	Decrease PTH incidence (Y/N)	Reduce pain (Y/N)	Study type	Commercial Products	Administer type	Other outcomes	FDA approval for human (Y/N)
Antifibrinolytics tranexamic acid (TXA) liquid	Y [83, 84]	N [83, 84]	–	Clinical reports analysis of 246 patients topically treated with TA and 248 control [84] Systematic review [83]	Cyklokapron®	Postop	no device or site for infection, reduce intraoperative blood loss during Orthognathic Surgery, prevent postop hemorrhage after oral or dental surgery in hemophilia A patients and those on warfarin anticoagulation [85]	N for Topical application
Natural material Propolis	–	Y [86]	Y [86]	Randomized controlled study on 65 patients [86]	Topical propolis gel	gargle immediately after surgery [86]	accelerated wound healing of tonsillar fossae	N
Herbal ingredients	Y [87]	–	–	Clinical trial in 47 consecutive children patients	Ankaferd Blood Stopper®	Intra-op	safe and efficient, and it reduces operating time [87], and contains no synthetic additives [88]	N
Autologous serum	–	Y [89]	Y [89]	A preliminary study on 32 patients (4–15 years old) [89]	Centrifuge peripheral venous blood at 1500–2000 g for 10 min to separate the serum and then topically administered	Intra and post 8 and 24 h	Contributed to tonsillar fossa epithelization in postoperative period Only preliminary study was performed	N

Table 2 Alternative biomaterials applied in other hemorrhage applications with potential for PTH

Biomaterial	Commercial Product	Mechanism of action	Application	Advantages	Drawbacks	Action/ resorption time	FDA approval (Y/N)
Scaffold/ Matrix Agents	Gelfoam®, Surgifoam®	porous matrix for platelet adhesion and fibrin clot formation; increased volume can provide hemostasis [29]	otosurgery [97], cardiac [63], lumbar spine [98]	little allergic activity [29]	Increased granuloma and infection [29]	4–6 weeks	Y [99, 100]
	Colgel®, Helitene®, Avitene®, CollaPlug®, CollaTape®	bovine collagen provides a substrate that activates platelets and allows them to adhere, facilitate clot formation through the intrinsic coagulation pathway [29, 101]	Tonsil bleeding [102], epistaxis [103], cardiac surgery [104, 105]	does not swell [29]	Adhesions, foreign body reactions, or allergic reactions have been reported. Most often with Avitene [106].	8 weeks	Y [107–109] except Colgel
	CoStasis®	combination of bovine collagen and bovine thrombin in a calcium chloride buffer and autologous obtained plasma that are mixed in equal volumes and administered intraoperatively. Fibrinogen from the patient's plasma is cleaved by the thrombin in the collagen matrix	hepatic orthopedic cardiothoracic, and general surgical procedures [110, 111]	acceptable biocompatibility [112]	Low antigenicity of collagen and possible allergic reactions [113]	achieve hemostasis within 3 min in cardiac surgery [110], 10 min for general surgeries, hepatic and iliac crest surgery [111]	Y [114]
	Arista AH®, HemoStase MPH®	flowable potato starch powder engineered to dehydrate blood, enhance clotting, concentrate erythrocytes and platelets for more effective thrombus formation [29, 115]	cardiothoracic surgical procedures [116], cerebral hemostasis [117], nephrectomies [118], sinus [119], cardiac [110]	nonpyogenic, good safety record [29]	short resorption, limited efficacy compared to procoagulant hemostatic [29]	24–48 h.	Y [120]
Biologic hemostats	Bovine origin: Thrombin-JM®, Thrombogen™ Human pooled: Evithrom®, Recombinant Human (rh): Recothrom®	Thrombin initiates cleavage of fibrinogen to fibrin, promoting clot formation [29]	comparative trial in liver resection, spine, peripheral arterial bypass, and dialysis access surgery [121]	RhThrombin reduced bleeding and thrombotic complications and less immunogenic than bovine thrombin [121]	Activation of autoimmune antibodies, causing profound coagulopathy [122], infectious disease concerns [29]	immediate	N bovine origin thrombin Y Evithrom [123] Y Recothrom [124]
	Spin blood at 3000 rpm for 10 min to get fibrin clot: the middle layer between the red corpuscles at bottom	Platelet cytokines, growth factors, and cells are entrapped and discharged in fibrin meshwork serving	hemostatic material for the treatment of oral lesions [126–128]	Cost-effective, simplified process, minimum blood manipulation and immunological reaction, promotes soft tissue healing	Need proper protocol and quick handling	–	N

Table 2 Alternative biomaterials applied in other hemorrhage applications with potential for PTH (Continued)

Biomaterial	Commercial Product	Mechanism of action	Application	Advantages	Drawbacks	Action/ resorption time	FDA approval (Y/N)
Chitin/chitosan dressings	ChitoFlex®, HemCon®, Celox™	<p>as a resorbable film [125], and platelets trigger blood clot and wound healing [125].</p> <p>Promote local vasoconstriction and serve as a scaffold for erythrocyte agglutination; physically occlude the wound site; stimulate fibroblast activation and collagen deposition [86]. Interacts with red blood cells and platelets directly to form a clot independent of clotting factors [110].</p>	<p>Cardiac [110], common homeostatic dressing for US military and pre-hospital wound dressing. [110], dental [129]</p>	<p>Antimicrobial due to acidic PH [130], efficacy in acquiring hemostasis and promoted initial healing phases [29, 110], nonallergic, nonexothermic, low cost, able to function in a hypothermic environment [110]</p>	<p>requires dressing and substrate to be in contact with wound</p>	<p>immediate to complete wound healing</p>	<p>Y HemoCon® [131, 132] Y Celox™ [131, 133]</p>
Hemocoagulase liquid	Hemocoagulase agkistrodon	<p>pharmacologic combination of coagulative proteins present in the venom of <i>Bothrops jararaca</i> or <i>Bothrops atrox</i> that works by directly cleaving fibrinogen to fibrin and activating factor X. Clot is thus formed independent of thrombin and will not break down in response to antithrombin [29, 134]</p>	<p>minor oral surgical procedures (impactions, simple extractions, transalveolar extractions) [135], postextraction bleeding [136]</p>	<p>Proven to significantly decrease postextraction bleeding [29]</p>	<p>Limitation in purchase and preparation</p>	<p>immediate</p>	<p>N</p>
granular mineral zeolite-based hemostatic agent (powder in a gauze mesh)	QuikClot®	<p>Zeolite absorbs water and concentrates coagulation factor, causes local dehydration with concentration of erythrocytes and platelets as well as activation of factor XII to initiate the coagulation cascade [29, 110, 137]</p>	<p>Cardiac surgeries [110], exsanguinating extremity wound and bleeding [131]</p>	<p>Cost-effective, stable</p>	<p>cannot be left in the wound site due to a foreign body reaction occurring</p>	<p>immediate</p>	<p>Y [131, 138]</p>
alginate dressing	Silverlon™ Antimicrobial Calcium Alginate Dressing	<p>Dehydration and concentration of erythrocytes and platelets, cationic initiation of coagulation cascade,</p>	<p>effective barrier to microbial penetration for moderate to heavy exuding wounds, and other surgical</p>	<p>Able to be removed with water without disrupting the underlying tissue healing</p>	<p>Not effective to handle a high flow of blood</p>	<p>immediate to complete wound healing</p>	<p>Y [141]</p>

Table 2 Alternative biomaterials applied in other hemorrhage applications with potential for PTH (Continued)

Biomaterial	Commercial Product	Mechanism of action	Application	Advantages	Drawbacks	Action/ resorption time	FDA approval (Y/N)
Tissue Adhesives							
Cyanoacrylates	Omnex®	barrier protection [139] The sealant polymerizes to form a flexible sealing film, which is adherent to both synthetic material and human tissue in a process that is independent of the patient's clotting processes.	wounds [140] Cardiothoracic [142], Neurosurgery [143]	Safe, strong, non-toxic, flexible, biocompatible, prevented blood leakage along suture, diminished hemostasis in coagulopathy, reduced time to hemostasis and seal	if closed in a wound, can lead to tissue necrosis and local inflammation; also do not breaking down into smaller absorbable fragments.	immediate to wound healing. The seal degrades with time, breaking down into smaller absorbable fragments.	Y [144]
Polyethylene glycol hydrogel	CoSeal®, DuraSeal®	2-phase application that results in the formation of a hydrogel matrix that becomes cross-linked with local proteins such as collagen [110]	Cardiac [145], lung [146], laparoscopic lymphadenectomy [147], urologic surgery	Found to be effective for urologic and vascular surgery and for CSF leak [29]	swells to 4 times its size	immediate	Y [148, 149]
Albumin-based–bovine-derived albumin cross-linked by glutaraldehyde	BioGlue®	Mixture of albumin-based-bovine-derived albumin with glycerinaldehyde creates strong crosslinks generating a tough hemostatic and adhesive matrix [29]	Cardiac [132], laparoscopic nephrectomy [150], secure hemostasis at cardiovascular anastomoses [151]	hemostatic and barrier protection	hypersensitivity reaction [29], impairs aortic growth [151], nerve tissue injury, cannot be used in pediatric cases as it impairs tissue growth [110].	immediate/indeterminate 20–30 s and reaches bonding strength by 2 min [110]	Y [152]
Mucosal tissue dressing based on methyl cellulose [153]	US9381270B2Acclarent, Inc., Menlo Park, CA (US)	Certain embodiments provide a biodegradable film or covering that serves as a mechanical barrier to reduce pain caused	for reducing or eliminating pain after Surgical procedures related to mucosal tissue tonsillectomy, adenoidectomy, or other pharyngeal operations.	Reduce pain and bleeding, facilitate mucosal tissue healing	Not clinical tested yet	Expected to be dissolved at 14 days and stable for 5 days	N

Table 2 Alternative biomaterials applied in other hemorrhage applications with potential for PTH (Continued)

Biomaterial	Commercial Product	Mechanism of action	Application	Advantages	Drawbacks	Action/ resorption time	FDA approval (Y/N)
Natural polymer based tissue adhesive (polysaccharides or partial hydrolysis derivatives or neutralization salts, chitosan and an alginate, carboxylic acid (acetic acid and lactic acid)) [154]	US20190038798A1 Ronnie Michael Hanes, Union Grove, AL (US); Adele Lamping Hanes, Union Grove, AL (US)	intraoperatively applied on the tonsil fossa which is then closed with the adhesive or sutures; used postoperatively as external dressing by application as a gel, thin film device or dry powder, or any methods in combination.	Post – operative application for tonsillectomy or adenoidectomy surgery, internal tissue adhesive for surgery or wound repair, application to a burn or skin donor site. For internal use, an optional treatment to improve resistance of the activated adhesive to body fluids is also described .	Promote healing with enhanced adhesive properties	Not fully tested yet	-	N

which was reported in association with the use of absorbable gelatin-based hemostats in nasal surgery [155]. Though there were no adverse effects of GTHM reported in secondary PTH [61], it should be proved consistently beneficial effect on pain and hemorrhage to be used for PTH.

Oxidized cellulose-based biomaterial

Oxidized cellulose-based biomaterials have been used for decades for their hemostatic properties in the control of oozing from broad surfaces. They are prepared by the oxidation of cellulose with nitrogen tetroxides (N_2O_4) [156]. Oxidized cellulose-based biomaterials contain no animal or human components, and are fabricated into mesh, gauze, woven strips, fibrillary tufts, and sponges. Oxidized cellulose acts as a hemostatic by providing a physical matrix for blood absorption and platelets adhesion and aggregation [157]. This accelerates the formation of a platelet plug and fibrin clots, improving wound healing and hemostasis. The low pH of the cellulosic acid within the biomaterial contributes to hemostasis by causing localized vasoconstriction and initial denaturation of blood proteins. The caustic pH exhibits immediate antibacterial effects, minimizing the risk of infection [158], while bacteriostatic and bactericidal properties inhibit growth of gram-positive and gram-negative organisms [159]. Additionally, cellulose-based biomaterials are entirely absorbable. The absorption of cellulose-based biomaterial takes 1 week to 4 weeks, depending on the products [113]. For tonsillectomy, commercial products need to be selected for specific application. For example, GuraTamp® can be completely absorbed within 7–14 days and provide a better option than Gelita-Cel® which can be completely biodegraded in 4 weeks.

Commercial absorbable hemostats products include Cellistyp® (B. Braun Medical Ltd., UK), Pahacel® (Effebe Hospital, Italy), Oxycel® (Becton Dickinson, Franklin Lakes, NJ), Gelita® (Gelita Medical GmbH, Germany), GuraTamp® (Jorgensen Lab, Loveland, CO), and Surgicel® (Ethicon). Surgicel is the most used cellulose-based hemostat in the United States. It is a fibrillary, oxidized-cellulose material in a sterile fabric meshwork. It has been widely used in otolaryngology and oral and maxillofacial surgery in order to control intra-osseous hemorrhage [73]. Its related products have had premarket approval from the FDA since 1960 [77]. A review of topical hemostatic agents in 2010 found that Surgicel had been used in otolaryngologic procedures, such as mastoid and endonasal surgeries [60]. For tonsillectomy, Surgicel application plus suturing decreases PTH rates [74].

No oxidized cellulose-based biomaterial products demonstrate strong adhesive properties, leading to a risk of aspiration and suffocation [75], as well as a potential

risk of inflammation [76]. To reduce the risk in tonsillectomy, cellulose-based biomaterial must be combined with suturing which can prolong operative time and adversely impact outcomes [74].

Bismuth subgallate-based materials

Bismuth subgallate (BSG) is an insoluble compound that has been showed with conflicting results in reducing hemorrhage incidence after tonsillectomy. The mechanism by which BSG achieves hemostasis is by accelerating the intrinsic clotting pathway via activation of Hageman factor XII [82]. After tonsillectomy performed by dissection and ligation without Diathermy, BSG mixed with adrenaline showed a significant decrease in hemostasis time and operating time in 39 patients, compared with 33 patients who did not receive BSG paste [81]. A prospective randomized trial of 60 patients showed that BSG/adrenaline paste reduced operative blood loss and operative time significantly [78]. One review of medical records showed that BSG reduces the incidence of primary tonsillar hemorrhage [80]. The use of BSG and epinephrine mixture has been reported with a low post-operative bleeding rate (4 of 1428 cases) [79]. The BSG-based paste was shown to be a faster and safer hemostat for PTH compared with the average pediatric tonsillectomy without BSG treatment [160, 161]. However, other studies suggest that BSG did not reduce hemorrhage [162, 163] or post-operative morbidity [81]. In the United States, BSG is an active ingredient in Devrom® (internal deodorant), an over-the-counter FDA-approved medicine [62]. The side effects of BSG include temporary darkening of the tongue and risk of foreign body response that may result in acute pneumonia [157, 164]. Due to BSG's possible hypersensitivity to the substance, hepatic or renal impairment is the caution impeding its application in patients with liver or kidney disease [157].

Antifibrinolytics

Antifibrinolytics are synthetic derivatives of the amino acid lysine. They inhibit fibrinolysis through binding to the enzyme plasmin and preventing the conversion of plasminogen to plasmin on the surface of the fibrin [32]. This prevents the lysis of the fibrin clot, which facilitates hemostasis and wound healing. Antifibrinolytics such as bovine aprotinin and TXA were used as additives in other hemostats for clot stabilization [155]. TXA has been proven effective for PTH by intravenous injection and oral medications, but less effective when applied topically [83, 165, 166].

Currently, TXA has been studied as a topical application for PTH and showed promising results in reducing blood loss in PTH, but no significant effect on PTH rate [83, 84]. CYKLOKAPRON® (Pfizer, New York City, NY) injection is a representative TXA commercial product,

which was approved by FDA in 1999 for intravenous injection but not for topical use [167]. TXA has been shown to be effective to prevent oral hemorrhage topically [168, 169]. Oral washes every 6 hours, starting peri-operatively and continuing for 7 days, have been used to prevent postoperative bleeding in patients with hemophilia A [170] and on oral anticoagulation [29, 168].

Side effects of TXA include thromboembolic risk in women who are using combination hormonal contraception, or those who have active or an intrinsic risk of thromboembolic disease. TXA has been implicated in cases venous and arterial thrombosis or thromboembolism, as well as cases of retinal artery and retinal vein occlusions [171]. Due to the life-threatening risk, studies on topical application of TXA in tonsillectomy must be developed to elucidate the benefit over intravenous injection for PTH. A large and well-designed, randomized and controlled trial is needed to investigate the risks and benefits in secondary PTH.

Natural materials

Propolis is a resinous mixture produced by mixing bee saliva and beeswax with exudate gathered from tree buds, sap flows, or other botanical sources. Propolis (Seoul Propolis Co., Korea) was immediately applied to bilateral tonsil fossae after surgery and then gargled by patients in a randomized study. The results demonstrated the efficacy of propolis in preventing PTH (from 16.9 to 4.6%), reducing pain, and accelerating wound healing of tonsillar fossae [86]. However, propolis is not approved by the FDA yet and only one study of propolis could be found. To confirm the effect, more clinical trials need to be conducted.

Another natural material, Ankaferd Blood Stopper® (Trend Technology Ilac AS, Turkey), is fabricated with herbal ingredients (plant extracts prepared from *Alpinia officinarum*, *Glycyrrhiza glabra*, *Thymus vulgaris*, *Urtica dioica* and *Vitis vinifera*). It was tested in 47 pediatric patients undergoing tonsillectomy by applying Ankaferd blood stopper on right tonsillar fossa. Patient's served as their own control with the left tonsillar fossa being knotted. Compared to the left, the right side was found significant less bleeding (1.57 ± 2.26 ml vs. 14.04 ± 7.23 ml) and operating time (3.19 ± 0.74 min vs. 7.29 ± 2.33 min) [87]. It is safe and effective, and contains no synthetic additives [88]. It has also showed to improve hemostasis in patients with acute anterior epistaxis [172].

Topically administered autologous serum was found contributed to reduce throat pain and hemorrhage rate in 32 post-tonsillectomy patients aged 4 to 15. In this preliminary study, autologous serum was prepared by centrifuging patient's blood at 1500–2000 g for 10 min. Then it was administered topically to the right tonsillar fossa for 10 min during the operation and at 8, 24 h

post-operatively. It was proposed that the hemostatic benefits were driven by epitheliotropic factors in the autologous serum speeding up the healing process [89]. Additionally, the serum has antibacterial properties due to the immunoglobulin G and lysozymes present therein. It has been used for ocular surface disease treatment and intra-articular and intraosseous injections [173, 174].

However, these natural materials have not been adequately studied in the context of PTH. Their influence on PTH severity, morbidity, and side effect profile need to be verified with more clinical studies.

Alternative biomaterials with potential for PTH

In addition to the aforementioned biomaterials which have been directly studied in PTH prevention, there are countless more biomaterials for achieving hemostasis intraoperatively that merit consideration for PTH prevention. Of the alternative biomaterials, those that could prove useful in preventing PTH can be categorized as scaffold agents, biologic hemostats, or tissue adhesives (Table 2).

Scaffold agents

Microporous mucopolysaccharide spheres (MMS) are small, porous spheres of potato starch whose pores serve as small sieves, wicking away liquid from the wound site and concentrating coagulation factors [29]. The product is a powder which is applied intraoperatively and is resorbed in approximately 24 to 48 h [29]. Arista Ah Absorbable Hemostat® (Medafor Inc., Minneapolis, MN) and HemoStase MPH® (Cryolife Inc., Atlanta, GA) are two commercially available products which have been used in cardiac, orthopedic, spinal, and general surgeries [110]. These products using licensed Microporous polysaccharide hemispheres (MPH) technology received premarket FDA approval [120]. They have been shown to be effective in reducing postoperative bleeding in cardiothoracic surgery [116] and endoscopic sinus surgery without negative impact on nasal mucosa healing, pain, and obstruction in the latter [175, 176]. Ease of use, low risk profile, and compatibility with mucosal healing all make MMSs exciting for PTH prevention, but their short duration of action and inferior hemostatic properties represent significant drawbacks.

Some scaffold products belong to gelatin-based biomaterials and act via the same mechanism as hemostats mentioned in section 1. These products include Gelfoam® (Pfizer) and Surgifoam® (Ethicon) (Table 2), and they, and their related products, have been approved by the FDA [99, 100]. Gelfoam is a water-insoluble, off-white, nonelastic, porous, pliable product prepared from purified porcine skin gelatin. The matrix is fully hydrolyzed by approximately 4 to 6 weeks [29]. It has been widely used to stop hemorrhage in surgeries such as

otosurgery [97], cardiac [63], and lumbar spinal surgery [98]. The hemostatic properties of gelatin matrices, their durability in the oral cavity, and their relatively low risk profile make them a potentially promising material for PTH prevention.

Microfibrillar collagen also shows promise as an easy-to-use strategy for reducing PTH. Microfibrillar collagen is a bovine collagen product that can be produced in powders, sheets, or plugs. It provides a substrate for platelet adherence and activation, thereby facilitating the formation of the fibrin clot [177]. It takes approximately 8 weeks for full resorption of the material [29]. Commercial products are Colgel® (Laboratoire Interphar, Aubervilliers, France), Helitene® (Integra Lifesciences Corporation, Plainsboro, NJ), Avitene® (Bard Davol, Warwick, RI), CollaPlug® and CollaTape® (Zimmer, Warsaw, IN). Colgel has been shown to reduce postoperative bleeding in cardiac surgery [104], while CollaTape and CollaPlug have been used to achieve hemostasis in oral extractions, grafts, and wound sites [178]. These products—except Colgel®—were approved by FDA [107–109]. Microfibrillar collagen has been reported to cause granuloma formation at the site of use [179], and prion disease is always a theoretical concern with the use of bovine products [110].

CoStasis® (Cohesion Technologies, Palo Alto, CA) is another formulation of microfibrillar collagen, composed of bovine thrombin and bovine collagen matrix. Intraoperatively, equal parts of CoStasis and autologous plasma are combined and administered as a sprayable liquid, creating a substrate loaded with thrombin which then cleaves the patient's fibrinogen, leading to clot formation [111]. It showed improved hemostasis as compared to manual compression in orthopedic, general, hepatic, and cardiac operations [111]. After a clinical study comparing 167 patients treated with CoStasis and 151 controls showed increased efficacy of CoStasis in obtaining hemostasis over standard methods, it was recommended to be used in difficult-to-manage hemorrhage [111].

Biological hemostats

Topical thrombin liquid (spray) including bovine origin and human origin have been tested as hemostats biomaterial. Recombinant human thrombin is a promising candidate because it reduced bleeding and thrombotic complications while being less immunogenic than bovine thrombin [121]. Bovine thrombin products such as Thrombin-JMI® (Pfizer) and Thrombogen™ (Johnson & Johnson) are not approved by FDA because of the immunogenic effects in human. Human thrombin products approved by FDA include Evithrom® (OMRIX Biopharmaceuticals, Inc., New York City, NY) and Recothrom® (ZymoGenetics, Inc. Seattle, WA) [123, 124]. The application of thrombin is usually combined with other

substrates, such as a damp sponge or gelatin sponge, to apply a thrombin solution to the wound site. It has been shown to be an effective hemostat in comparative trials in liver resection, spinal surgery, peripheral arterial bypass, and dialysis access surgery [121]. However, pooled human thrombin raises infectious disease concerns [29], while purified bovine thrombin activates autoimmune antibodies—typically against factor V—potentially causing profound coagulopathy [122].

More recently (2018 and 2019), it was found that the use of platelet-rich fibrin membranes may represent a feasible alternative hemostatic material for the treatment of oral lesions [126, 180]. Platelets acts as autologous sources of cytokines and growth factors to treat hemorrhage. Platelet concentrate derived from blood can be used to prevent and treat bleeding due to severe thrombocytopenia and oral hemorrhage associated with medullary aplasia, acute leukemia [181]. Because of the role of platelets in coagulation, platelet-rich membranes might have a promising application in post-tonsillectomy hemorrhage.

Chitin is a natural substance found in the exoskeletons of arthropods, the cell walls of some fungi, or as a byproduct of algae fermentation. Chitin and its derivative chitosan have myriad beneficial effects on hemostasis and wound healing. Chitin and chitosan facilitate hemostasis by providing a scaffold for erythrocyte aggregation, increasing platelet and clotting factor delivery to the wound, and by promoting local vasoconstriction [29, 178]. Further, both have been shown to improve the speed of wound healing and reconstruction of connective tissue, in addition to having painkilling and antimicrobial properties [182–186]. Formulations are gels, fibers, films, or beads, with many options for application in the oral cavity [29]. Biodegradation studies have shown persistence of chitosan at wound sites up to 14 days after administration, with minimal local or systemic risks [187]. ChitoFlex® and HemCon® (HemCon Medical Technologies Inc., Portland, OR), Celox™ (SAM Medical Products, Portland, OR) are commercial chitin/chitosan products. Novel chitosan delivery methods, such as polyvinylpyrrolidone-chitosan film and chitosan-alginate hydrogel, are in development to be applicable in the oral cavity [186, 188]. Chitin and chitosan represent one of the most promising materials that has yet to be directly studied in PTH prevention.

Additional biomaterials listed in Table 2 include Hemocoagulase agkistrodon (Konruns Pharmaceutical Co., Ltd., China), zeolite-based QuikClot® (Z-Medica Corp, Wallingford, CT) and alginate dressing Silverlon™ Antimicrobial Calcium Alginate Dressing (Argentum Medical, LLC, Geneva, IL). They have been applied in oral and cardiac surgeries, and proved effective in hemostasis and wound healing. But each has limitations: profound coagulopathy by topical thrombin liquid,

handling difficulty of platelet-rich fibrin membranes, and lack of substrate for chitin/chitosan dressing. Addressing these limitations will promote an immediately apparent utility in PTH prevention.

Tissue adhesives

Tissue adhesives have been widely used to provide hemostasis for PTH (fibrin-based sealant in Table 1). Other types of tissue adhesives (Table 2) have been applied as hemostats in oral and maxillofacial surgeries [29]. The commercial products of the listed tissue adhesives are 1) Cyanoacrylates: Omnex® Surgical Sealant (Ethicon); 2) polyethylene glycol hydrogel: CoSeal® (Baxter International) and DuraSeal® (Integra lifesciences, Plainsboro, NJ); and 3) Mixture of bovine serum albumin (BSA) and glutaraldehyde: Bioglu® (CryoLife Inc., Atlanta, GA). Methyl cellulose based mucosal tissue dressing [153] and nature polymer based tissue adhesive [154] are patent describing desirable adhesive properties for reducing post-operative pain and bleeding, and promoting wound healing. Information regarding the application, advantages, drawbacks, and FDA approval status are listed in Table 2. Any biomaterial that would be used for PTH prevention would be exposed to consistent shear forces and a complex environment as soon as applied to the wound site in the oropharynx. The adhesive biomaterials provide a mechanical barrier to protect tonsil wound from friction by food in the first few days after surgery, thus enhance healing after tonsillectomy. Thus high adhesive strength is one of the most required properties for PTH that one biomaterial should be manufactured with.

Conclusion

The lack of widespread clinical adoption of both current and potential biomaterials is due to the following difficulties in PTH prevention: 1) The vascular supply to the tonsil is robust, making tonsillectomy the “ultimate test of hemostasis”; 2) The complex environment is routinely exposed to saliva and bacteria; 3) Shear forces from swallowing (food viscosity) and coughing (high flow rate) make tissue adhesion challenging. These three criteria can be used as an a priori assessment of the potential utility of any biomaterial in PTH prevention.

Any biomaterial that would be used in the prevention of PTH should be biocompatible, easy to apply, cost efficient, reduce complications, remain stable as protective physical barrier, and promote wound healing and remucosalization. Among the reviewed current biomaterials for PTH, the fibrin-based collagen fleece and gelatin-based patch are more effective for PTH than the other biomaterials. In alternative biomaterials, tissue adhesives and chitin/chitosan materials seem promising to be tested for PTH, because their adhesive properties might benefit hemostasis in the tonsil environment. The

most common mechanism of action utilized to prevent PTH is formation of the fibrin clot which improves wound healing and hemostasis. For the developing biomaterials to prevent PTH, it is necessary to perform comparative clinical trials to elucidate their capacities to achieve hemostasis, their biocompatibility, and their adhesive properties.

Abbreviations

BSA: Bovine serum albumin; BSG: Bismuth subgallate; FDA: Food and drug administration; GTHM: Gelatin-thrombin hemostatic matrix; MMS: Microporous mucopolysaccharide spheres; MPH: Microporous polysaccharide hemispheres; N2O4: Nitrogen tetroxides; PTH: Post-tonsillectomy hemorrhage; TXA: Tranexamic acid

Acknowledgements

The authors thank the support from all colleagues of Tissue-Engineering and Surgical Research Institute and Department of Pediatric Otorhinolaryngology at Nationwide Children's Hospital.

Authors' contributions

Conceptualization: TC. Manuscript configuration: LL. Writing original draft preparation: LL, CR. Writing review and editing: LL, CR, NW, JJ, CE, TC. All authors read and approved the final manuscript. No authors have any financial or non-financial competing interests in regards to this manuscript.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

Availability of data and materials

Not applicable.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Center of Regenerative Medicine, Research Institute at Nationwide Children's Hospital, Columbus, OH, USA. ²College of Medicine, The Ohio State University, Columbus, OH, USA. ³Department of Pediatric Otorhinolaryngology, Nationwide Children's Hospital, Columbus, OH, USA. ⁴Nanofiber Solutions, Hilliard, OH, USA.

Received: 20 May 2019 Accepted: 27 August 2019

Published online: 06 September 2019

References

- Clark CM, Schubart JR, Carr MM. Trends in the management of secondary post-tonsillectomy hemorrhage in children. *Int J Pediatr Otorhinolaryngol.* 2018;108:196–201.
- Cullen KA, Hall MJ, Golosinskiy A. Ambulatory surgery in the United States, 2006; 2009.
- Darrow DH, Siemens C. Indications for tonsillectomy and adenoidectomy. *Laryngoscope.* 2002;112(S100):6–10.
- Wall JJ, Tay K-Y. Postoperative tonsillectomy hemorrhage. *Emerg Med Clin.* 2018;36(2):415–26.
- Steketee KG, Reisdorff EJ. Emergency care for posttonsillectomy and postadenoidectomy hemorrhage. *Am J Emerg Med.* 1995;13(5):518–23.
- Windfuhr J, Schloendorff G, Baburi D, Kremer B. Serious post-tonsillectomy hemorrhage with and without lethal outcome in children and adolescents. *Int J Pediatr Otorhinolaryngol.* 2008;72(7):1029–40.
- Windfuhr J, Schloendorff G, Baburi D, Kremer B. Lethal outcome of post-tonsillectomy hemorrhage. *Eur Arch Otorhinolaryngol.* 2008;265(12):1527.

8. Chowdhury K, Tewfik T, Schloss M. Post-tonsillectomy and adenoidectomy hemorrhage. *J Otolaryngol*. 1988;17(1):46–9.
9. Irani DB, Berkowitz RG. Management of secondary hemorrhage following pediatric adenotonsillectomy. *Int J Pediatr Otorhinolaryngol*. 1997;40(2–3):115–24.
10. Windfuhr JP. Excessive post-tonsillectomy hemorrhage requiring ligation of the external carotid artery. *Auris Nasus Larynx*. 2002;29(2):159–64.
11. Windfuhr JP, Chen Y-S. Incidence of post-tonsillectomy hemorrhage in children and adults: a study of 4,848 patients. *Ear Nose Throat J*. 2002;81(9):626–32.
12. Evans A, Khan A, Young D, Adamson R. Assessment of secondary haemorrhage rates following adult tonsillectomy—a telephone survey and literature review. *Clin Otolaryngol Allied Sci*. 2003;28(6):489–91.
13. Capper J, Randall C. Post-operative haemorrhage in tonsillectomy and adenoidectomy in children. *J Laryngol Otol*. 1984;98(4):363–5.
14. Peeters A, Claes J, Saldien V. Lethal complications after tonsillectomy. *Acta Otorhinolaryngol Belg*. 2001;55(3):207–13.
15. Densert HD, Eliasson A, Frederiksen L, Andersson O, Olaison J, Ove CW. Tonsillectomy in children with tonsillar hypertrophy. *Acta Otolaryngol*. 2001;121(7):854–8.
16. Linder A, Markström A, Hultcrantz E. Using the carbon dioxide laser for tonsillectomy in children. *Int J Pediatr Otorhinolaryngol*. 1999;50(1):31–6.
17. Moriniere S, Roux A, Bakhos D, Trijolet J-P, Pondaven S, Pinlong E, et al. Radiofrequency tonsillectomy versus bipolar scissors tonsillectomy for the treatment of OSAS in children: a prospective study. *Eur Ann Otorhinolaryngol Head Neck Dis*. 2013;130(2):67–72.
18. Çelenk F, Bayazit YA, Yılmaz M, Kemaloglu YK, Uygur K, Ceylan A, et al. Tonsillar regrowth following partial tonsillectomy with radiofrequency. *Int J Pediatr Otorhinolaryngol*. 2008;72(1):19–22.
19. Ripplinger T, Theuerkauf T, Schultz-Coulon H. Significance of the medical history in decisions on whether tonsillectomy is indicated. *HNO*. 2007;55(12):945–9.
20. Dalesio NM. Management of post-tonsillectomy hemorrhage. *Cases Emerg Airway Manage*. 2015;24:184–9.
21. Davidoss N, Eikelboom R, Friedland P, Santa MP. Wound healing after tonsillectomy—a review of the literature. *J Laryngol Otol*. 2018;132(9):764–70.
22. McGovern FH. Prevention of secondary post-tonsillectomy hemorrhage with sulfathiazole gum. *Arch Otolaryngol*. 1944;40(3):196–7.
23. Ferguson J, Nürnberger S, Redl H. Fibrin: the very first biomimetic glue—still a great tool. *Biological adhesive systems*: Springer. 2010;225–36.
24. Janmey PA, Winer JP, Weisel JW. Fibrin gels and their clinical and bioengineering applications. *J R Soc Interface*. 2008;6(30):1–10.
25. Quinn JV. *Tissue adhesives in clinical medicine*: PMPH-USA; 2005.
26. Nürnberger S, Wolbank S, Peterbauer-Scherb A, Morton TJ, Feichtinger GA, Gugerell A, et al. Properties and potential alternative applications of fibrin glue. *Biological adhesive systems*: Springer. 2010;237–59.
27. Rosso F, Marino G, Giordano A, Barbarisi M, Parmeggiani D, Barbarisi A. Smart materials as scaffolds for tissue engineering. *J Cell Physiol*. 2005;203(3):465–70.
28. Oz MC, Rondinone JF, Shargill NS. FloSeal matrix: new generation topical hemostatic sealant. *J Card Surg*. 2003;18(6):486–93.
29. Vezeau PJ. Topical hemostatic agents: what the oral and maxillofacial surgeon needs to know. *Oral Maxillofac Surg Clin*. 2016;28(4):523–32.
30. Hegab AA. Omentoplasty versus tachosil in preventing leakage after colonic anastomosis. *Egyptian J Surg*. 2016;35(4):372.
31. Sprout R, Radford P, Hunt A. Hemostatic glues in tonsillectomy: a systematic review. *Laryngoscope*. 2016;126(1):236–42.
32. Aronson JK. *Meyler's side effects of drugs* (16th Edition): the international encyclopedia of adverse drug reactions and interactions. Elsevier Science; 2015.
33. Tisseel®, Fibrin Sealant Patch, authors FDA Approval Letter. 1998.
34. Stoeckli SJ, Moe KS, Huber A, Schmid S. A prospective randomized double-blind trial of fibrin glue for pain and bleeding after tonsillectomy. *Laryngoscope*. 1999;109(4):652–5.
35. Segal N, Puterman M, Rotem E, Niv A, Kaplan D, Kraus M, et al. A prospective randomized double-blind trial of fibrin glue for reducing pain and bleeding after tonsillectomy. *Int J Pediatr Otorhinolaryngol*. 2008;72(4):469–73.
36. Vaiman M, Shlankovitch N, Eviatar E, Segal S. Effect of modern fibrin glue on bleeding after tonsillectomy and adenoidectomy. *Ann Otol Rhinol Laryngol*. 2003;112(5):410–4.
37. Stiller-Timor L, Goldbart A, Segal N, Amash A, Huleihel M, Leiberman A, et al. Circulating cytokines in patients undergoing tonsillectomy with fibrin glue. *Int J Pediatr Otorhinolaryngol*. 2012;76(3):419–22.
38. Vaiman M, Krakovski D, Gavriel H. Fibrin sealant reduces pain after tonsillectomy: prospective randomized study. *Ann Otol Rhinol Laryngol*. 2006;115(7):483–9.
39. Dhillon S. Fibrin sealant (Evicel®/Quixil®/Crosseal™). *Drugs*. 2011;71(14):1893–915.
40. Vaiman M, Eviatar E, Segal S. Effectiveness of second-generation fibrin glue in endonasal operations. *Otolaryngol Head Neck Surg*. 2002;126(4):388–91.
41. Vaiman M, Eviatar E, Segal S. The use of fibrin glue as hemostatic in endonasal operations: a prospective, randomized study. *Rhinology*. 2002;40(4):185–8.
42. Vaiman M, Segal S, Eviatar E. Fibrin glue treatment for epistaxis. *Rhinology*. 2002;40(2):88–91.
43. Richmon JD, Tian Y, Husseman J, Davidson TM. Use of a sprayed fibrin hemostatic sealant after laser therapy for hereditary hemorrhagic telangiectasia epistaxis. *Am J Rhinol*. 2007;21(2):187–91.
44. Brown AC, Barker TH. Fibrin-based biomaterials: modulation of macroscopic properties through rational design at the molecular level. *Acta Biomater*. 2014;10(4):1502–14.
45. Scheyer M, Zimmermann G. Tachocomb used in endoscopic surgery. *Surg Endosc*. 1996;10(5):501–3.
46. Nam JG, Lee T-H, Kwon JK, Lee JC, Lee SR, Lee SM, et al. Effect of fibrin-coated collagen fleece (TachoComb) on pain and bleeding after adenotonsillectomy in children. *Acta Otolaryngol*. 2011;131(12):1293–8.
47. Agger P, Langhoff J, Smerup MH, Hasenkam JM. Comparison between TachoComb and TachoSil for surgical hemostasis in arterial bleeding: an animal experimental study. *J Trauma Acute Care Surg*. 2010;68(4):838–42.
48. TachoSil®, authors FDA Approval Letter. 2010.
49. Yeoboh M. FDA approves first biodegradable sealant patch for cardiovascular surgery. FDA News Release. 2010. <https://www.fiercebiotech.com/biotech/fda-approves-first-biodegradable-sealant-patch-for-cardiovascular-surgery>.
50. Kawasaki S, Origasa H, Tetens V, Kobayashi M. Comparison of TachoSil and TachoComb in patients undergoing liver resection—a randomized, double-blind, non-inferiority trial. *Langenbecks Arch Surg*. 2017;402(4):591–8.
51. Berdajs D, Bürki M, Micheli A, von Segesser LK. Seal properties of TachoSil®: in vitro hemodynamic measurements. *Interact Cardiovasc Thorac Surg*. 2010;10(6):910–3.
52. Toro A, Mannino M, Reale G, Di Carlo I. TachoSil use in abdominal surgery: a review. *J Blood Med*. 2011;2:31.
53. Matonick JP, Hammond J. Hemostatic efficacy of EVARREST™, fibrin sealant patch vs. TachoSil® in a heparinized swine spleen incision model. *J Invest Surg*. 2014;27(6):360–5.
54. EVARREST® Fibrin Sealant Patch, authors FDA Approval Letter. 2012.
55. Koea JB, Batiller J, Aguirre N, Shen J, Kocharian R, Bochicchio G, et al. A multicentre, prospective, randomized, controlled trial comparing EVARREST™ fibrin sealant patch to standard of care in controlling bleeding following elective hepatectomy: anatomic versus non-anatomic resection. *HPB*. 2016;18(3):221–8.
56. Baker JE, Goodman MD, Makley AT, Stevens-Topie SM, Veile RA, Mahoney EJ, et al. Evaluation of a novel fibrin sealant patch in hemorrhage control after vascular or hepatic injury. *Mil Med*. 2018;184(3–4):290–6.
57. Gabay M, Boucher BA. An essential primer for understanding the role of topical hemostats, surgical sealants, and adhesives for maintaining hemostasis. *Pharmacother: J Human Pharmacol Drug Ther*. 2013;33(9):935–55.
58. Tărcoveanu E, Lupașcu C, Moldovanu R, Vlad N, Bradea C, Vasilescu A. Fibrin-collagen patch (TachoComb) in general surgery. Indications and results. *Rev Med Chir Soc Med Nat Iasi*. 2007;111(2):396–401.
59. Spotnitz WD. Fibrin sealant: the only approved hemostat, sealant, and adhesive—a laboratory and clinical perspective. *ISRN Surg*. 2014;2014.
60. Acar B, Babademez MA, Karabulut H. Topical hemostatic agents in otolaryngologic surgery. *Kulak Burun Bogaz Ihtis Derg*. 2010;20(2):100–9.
61. Binnetoglu A, Demir B, Yumusakhuyul AC, Baglam T, Sari M. Use of a gelatin-thrombin hemostatic matrix for secondary bleeding after pediatric tonsillectomy. *JAMA Otolaryngol Head Neck Surg*. 2016;142(10):954–8.
62. Bismuth Subgallate, Code of Federal Regulations, Drugs for human use, FDA. 2018;Title 21, Volume 5, Part 357, Subpart I (CITE: 21CFR357.850).
63. Oz MC, Cosgrove DM III, Badduke BR, Hill JD, Flannery MR, Palumbo R, et al. Controlled clinical trial of a novel hemostatic agent in cardiac surgery. *Ann Thorac Surg*. 2000;69(5):1376–82.

64. Pharmacy Co, Chemistry. Council on pharmacy and chemistry: absorbable gelatin sponge—new and nonofficial remedies. *JAMA*. 1947;135:921.
65. Spotnitz WD. Fibrin sealant: past, present, and future: a brief review. *World J Surg*. 2010;34(4):632–4.
66. Kim Y-W, Kang M-J, Lee H-J, Woo C-K, Mun M-J, Cho K-S. The efficacy of TachoComb on reducing postoperative complications after tonsillectomy in children. *Int J Pediatr Otorhinolaryngol*. 2015;79(8):1337–40.
67. Jo SH, Mathiasen RA, Gurushanthaiah D. Prospective, randomized, controlled trial of a hemostatic sealant in children undergoing adenotonsillectomy. *Otolaryngol Head Neck Surg*. 2007;137(3):454–8.
68. Blackmore K, O'Hara J, Flood L, Martin F. The effect of FloSeal on post-tonsillectomy pain: a randomised controlled pilot study. *Clin Otolaryngol*. 2008;33(3):281–4.
69. Mozet C, Prettin C, Dietze M, Fickweiler U, Dietz A. Use of FloSeal and effects on wound healing and pain in adults undergoing tonsillectomy: randomised comparison versus electrocautery. *Eur Arch Otorhinolaryngol*. 2012;269(10):2247–54.
70. Mathiasen RA, Cruz RM. Prospective, randomized, controlled clinical trial of a novel matrix hemostatic sealant in children undergoing adenoidectomy. *Otolaryngol Head Neck Surg*. 2004;131(5):601–5.
71. Echave M, Oyagüez I, Casado MA. Use of FloSeal®, a human gelatine-thrombin matrix sealant, in surgery: a systematic review. *BMC Surg*. 2014;14(1):111.
72. FloSeal™ Matrix, authors FDA Approval Letter. 1999.
73. Goodman RS. Surgicel in the control of post-tonsillectomy bleeding. *Laryngoscope*. 1996;106(8):1044–6.
74. Cetiner H, Cavusoglu I, Duzer S, Sakalloglu O, Susaman N, Yildirim YSS. Effect of Saturation plus Surgicel application on post-tonsillectomy bleeding and pain. *J Craniofac Surg*. 2017;28(7):e672–e5.
75. Ronen B, Itzhak B. The risks of Haemostatic materials in tonsillectomy. *Arch Otolaryngol Rhinol*. 2015;1(2):046–7.
76. Wang H, Chen P. Surgicel®(oxidized regenerated cellulose) granuloma mimicking local recurrent gastrointestinal stromal tumor: a case report. *Oncol Lett*. 2013;5(5):1497–500.
77. Surgicel®, Premarket Approval by FDA. PMA Number: N12159. 2018.
78. Agrawal S, Jain A, Marathe D, Agrawal R. The effect of bismuth subgallate as haemostatic agent in tonsillectomy. *Indian J Otolaryngol Head Neck Surg*. 2005;57(4):287–9.
79. Maniglia AJ, Kushner H, Cozzi L. Adenotonsillectomy: a safe outpatient procedure. *Arch Otolaryngol Head Neck Surg*. 1989;115(1):92–4.
80. Conley SF, Ellison MD. Avoidance of primary post-tonsillectomy hemorrhage in a teaching program. *Arch Otolaryngol Head Neck Surg*. 1999;125(3):330–3.
81. Fenton J, Blayney A, O'Dwyer T. Bismuth subgallate—its role in tonsillectomy. *J Laryngol Otol*. 1995;109(3):203–5.
82. Thorisdottir H, Ratnoff O, Maniglia A. Activation of Hageman factor (factor XII) by bismuth subgallate, a hemostatic agent. *J Lab Clin Med*. 1988;112(4):481–6.
83. Chan CC, Chan YY, Tanweer F. Systematic review and meta-analysis of the use of tranexamic acid in tonsillectomy. *Eur Arch Otorhinolaryngol*. 2013;270(2):735–48.
84. Hinder D, Tschopp K. Topical application of tranexamic acid to prevent post-tonsillectomy haemorrhage. *Laryngo-rhino-otologie*. 2015;94(2):86–90.
85. Eftekharian H, Vahedi R, Karagah T, Tabrizi R. Effect of tranexamic acid irrigation on perioperative blood loss during orthognathic surgery: a double-blind, randomized controlled clinical trial. *J Oral Maxillofac Surg*. 2015;73(1):129–33.
86. Moon JH, Lee MY, Chung Y-J, Rhee C-K, Lee SJ. Effect of topical Propolis on wound healing process after tonsillectomy: randomized controlled study. *Clin Experiment Otorhinolaryngol*. 2018;11(2):146.
87. Teker AM, Korkut AY, Gedikli O, Kahya V. Prospective, controlled clinical trial of Ankaferd blood stopper in children undergoing tonsillectomy. *Int J Pediatr Otorhinolaryngol*. 2009;73(12):1742–5.
88. Fisgin NT, Cayci YT, Coban AY, Ozatli D, Tanyel E, Durupinar B, et al. Antimicrobial activity of plant extract Ankaferd blood stopper®. *Fitoterapia*. 2009;80(1):48–50.
89. Kara M, Erdoğan H, Altinişik H, Aylanc H, Güçlü O, Dereköy F. Does topical use of autologous serum help to reduce post-tonsillectomy morbidity? A prospective, controlled preliminary study. *J Laryngol Otol*. 2016;130(7):662–8.
90. Renkens KL Jr, Payner TD, Leipzig TJ, Feuer H, Morone MA, Koers JM, et al. A multicenter, prospective, randomized trial evaluating a new hemostatic agent for spinal surgery. *Spine*. 2001;26(15):1645–50.
91. Ellegala DB, Maartens NF, Laws ER Jr. Use of FloSeal hemostatic sealant in transsphenoidal pituitary surgery. *Neurosurgery*. 2002;51(2):513–6.
92. Mathiasen RA, Cruz RM. Prospective, randomized, controlled clinical trial of a novel matrix hemostatic sealant in patients with acute anterior epistaxis. *Laryngoscope*. 2005;115(5):899–902.
93. Chandra RK, Conley DB, Haines GK III, Kern RC. Long-term effects of FloSeal™ packing after endoscopic sinus surgery. *Am J Rhinol*. 2005;19(3):240–3.
94. Gall RM, Witterick IJ, Shargill NS, Hawke M. Control of bleeding in endoscopic sinus surgery: use of a novel gelatin-based hemostatic agent. *J Otolaryngol*. 2002;31(5):271–4.
95. Giger B, Bonanomi A, Odermatt B, Ladell K, Speck RF, Kojic D, et al. Human tonsillar tissue block cultures differ from autologous tonsillar cell suspension cultures in lymphocyte subset activation and cytokine gene expression. *J Immunol Methods*. 2004;289(1–2):179–90.
96. Roy S, Bigcas J-L, Vandelaar L. Hemostasis in pediatric surgery. *Otolaryngol Clin North Am*. 2016;49(3):601–14.
97. Hellström S, Salén B, Stenfors L-E. Absorbable gelatin sponge (Gelfoam) in Otolaryngology: one cause of undesirable postoperative results?: an experimental study in the rat. *Acta Otolaryngol*. 1983;96(3–4):269–75.
98. Friedman J, Whitecloud TS III. Lumbar cauda equina syndrome associated with the use of gelfoam: case report. *Spine*. 2001;26(20):E485–E7.
99. Gelfoam®, Premarket Approval by FDA. PMA Number: N18286. 2018.
100. Surgifoam®, Premarket Approval by FDA. PMA Number: N990004. 2019.
101. Parker RK, Dinehart SM. Hints for hemostasis. *Dermatol Clin*. 1994;12(3):601–6.
102. Levy S, Brodsky L, Stanievich J. Hemorrhagic tonsillitis. *Laryngoscope*. 1989;99(1):15–8.
103. Walike J, Chinn J. Evaluation and treatment of acute bleeding from the head and neck. *Otolaryngol Clin North Am*. 1979;12(2):455–64.
104. Sirlak M, Eryilmaz S, Yazicioglu L, Kiziltepe U, Eyiletlen Z, Durdu MS, et al. Comparative study of microfibrillar collagen hemostat (Colgel) and oxidized cellulose (Surgicel) in high transfusion-risk cardiac surgery. *J Thorac Cardiovasc Surg*. 2003;126(3):666–70.
105. Robicsek F. Microfibrillar collagen hemostat in cardiac surgery. *J Thorac Cardiovasc Surg*. 2004;127(4):1228.
106. Alexander JM, Rabinowitz J. Microfibrillar collagen (Avitene) as a hemostatic agent in experimental oral wounds. *J Oral Surg*. 1978;36(3):202–5.
107. Helistat, Helitene Absorbable Collagen Hemostatic Agents®, Premarket Approval by FDA. PMA Number: P850010. 2018.
108. Avitene Microfibrillar Collagen Hemostat®, Premarket Approval by FDA. PMA Number: N17600. 2018.
109. Collacote collatape collaplyg absorbable collagen wound dressing®, Premarket Approval by FDA. PMA Number: P840062. 2019.
110. Barnard J, Millner R. A review of topical hemostatic agents for use in cardiac surgery. *Ann Thorac Surg*. 2009;88(4):1377–83.
111. Committee CM-cCW. A novel collagen-based composite offers effective hemostasis for multiple surgical indications: results of a randomized controlled trial. *Surgery*. 2001;129(4):445–50.
112. Buncke GM, Sherman R. Costasis® provides superior control of diffuse bleeding at muscle-flap donor sites, compared to manual compression. *J Reconstr Microsurg*. 2000;16(07):0557–62.
113. Tomizawa Y. Clinical benefits and risk analysis of topical hemostats: a review. *J Artif Organs*. 2005;8(3):137–42.
114. Costasis Surgical Hemostat®, Premarket Approval by FDA. PMA Number: P990030. 2002.
115. Tan SR, Tope WD. Effectiveness of microporous polysaccharide hemospheres for achieving hemostasis in Mohs micrographic surgery. *Dermatol Surg*. 2004;30(6):908–14.
116. Bruckner BA, Blau LN, Rodriguez L, Suarez EE, Ngo UQ, Reardon MJ, et al. Microporous polysaccharide hemisphere absorbable hemostat use in cardiothoracic surgical procedures. *J Cardiothorac Surg*. 2014;9(1):134.
117. Galarza M, Porcar OP, Gazzeri R, Martínez-Lage JF. Microporous polysaccharide hemospheres (MPH) for cerebral hemostasis: a preliminary report. *World Neurosurg*. 2011;75(3–4):491–4.
118. Rajagopal P, Hakim N. The use of a powdered polysaccharide hemostat (HemoStase) in live donor nephrectomies controls bleeding and reduces postoperative complications. *Transplantation proceedings: Elsevier Science*. 2011;43(2):424–6.
119. Beyea JA, Rotenberg BW. Comparison of purified plant polysaccharide (HemoStase) versus gelatin-thrombin matrix (FloSeal) in controlling bleeding during sinus surgery: a randomized controlled trial. *Ann Otol Rhinol Laryngol*. 2011;120(8):495–8.

120. Arista Ah absorbable Hemostat®, Premarket Approval by FDA. PMA Number: P050038. 2019.
121. Chapman WC, Singla N, Genyk Y, McNeil JW, Renkens KL Jr, Reynolds TC, et al. A phase 3, randomized, double-blind comparative study of the efficacy and safety of topical recombinant human thrombin and bovine thrombin in surgical hemostasis. *J Am Coll Surg*. 2007;205(2):256–65.
122. Lawson JH, Lynn KA, Vanmatre RM, Domzalski T, Klemp KF, Ortel TL, et al. Antihuman factor V antibodies after use of relatively pure bovine thrombin. *Ann Thorac Surg*. 2005;79(3):1037–8.
123. EVITHROM® Thrombin, Topical (Human), FDA Approval Letter, STN: BL 125247. 2007.
124. RECOThROM® Thrombin, Topical ((Recombinant), FDA Approval Letter, STN: BL 125248. 2008.
125. Gaßling VL, Açil Y, Springer IN, Hubert N, Wiltfang J. Platelet-rich plasma and platelet-rich fibrin in human cell culture. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2009;108(1):48–55.
126. CfDAB M, Calasans-Maia MD, de Mello Machado RC, de Brito Resende RF, Alves GG. The use of platelet-rich fibrin as a hemostatic material in oral soft tissues. *Oral Maxillofac Surg*. 2018;22(3):329–33.
127. Naik B, Karunakar P, Jayadev M, Marshal VR. Role of platelet rich fibrin in wound healing: a critical review. *J Conserv Dent*. 2013;16(4):284.
128. Kumar KR, Genmorgan K, Rahman SA, Rajan MA, Kumar TA, Prasad VS. Role of plasma-rich fibrin in oral surgery. *J Pharm Bioallied Sci*. 2016; 8(Suppl 1):S36.
129. McDevitt JP, Malik S. Hemostat coated dental floss and hemostat coated dental tape. Google Patents: US6536448B2. 2003.
130. Chan MW, Schwartzberg SD, Demcheva M, Vournakis J, Finkielstein S, Connolly RJ. Comparison of poly-N-acetyl glucosamine (P-GlcNAc) with absorbable collagen (Actifoam), and fibrin sealant (Bolheal) for achieving hemostasis in a swine model of splenic hemorrhage. *J Trauma Acute Care Surg*. 2000;48(3):454–8.
131. Devlin JJ, Kircher S, Kozen BG, Littlejohn LF, Johnson AS. Comparison of ChitoFlex®, CELOX™, and QuikClot® in control of hemorrhage. *J Emerg Med*. 2011;41(3):237–45.
132. Chao HH, Torchiana DF. BioGlue: albumin/glutaraldehyde sealant in cardiac surgery. *J Card Surg*. 2003;18(6):500–3.
133. CELOX TRAUMA GAUZE AG, CELOX HEMOSTATIC ANTIBACTERIAL TRAUMA GAUZE, OMNI-STAT TRAUMA GAUZE AG, OMNI-STAT HEMOSTATIC ANTI. 510 (k) Premarket Notification, FDA, K102965. 2010.
134. Shenoy AK, Ramesh K, Chowta MN, Adhikari PM, Rathnakar U. Effects of botropase on clotting factors in healthy human volunteers. *Perspect Clin Res*. 2014;5(2):71.
135. Majumder K, Rao SJD, Gehlot N, Arya V, Siwach V. Efficacy of Haemocoagulase as a topical Haemostatic agent after minor Oral surgical procedures—a prospective study. *Int J Clin Med*. 2014;5(14):875.
136. Joshi SA, Gadre KS, Halli R, Shandilya R. Topical use of Hemocoagulase (Reptilase): a simple and effective way of managing post-extraction bleeding. *Ann Maxillofacial Surg*. 2014;4(1):119.
137. Griffin JH. Role of surface in surface-dependent activation of Hageman factor (blood coagulation factor XII). *Proc Natl Acad Sci*. 1978;75(4): 1998–2002.
138. Li J, Cao W, Lv XX, Jiang L, Li YJ, Li WZ, et al. Zeolite-based hemostat QuikClot releases calcium into blood and promotes blood coagulation in vitro. *Acta Pharmacol Sin*. 2013;34(3):367.
139. Segal HC, Hunt BJ, Gilding K. The effects of alginate and non-alginate wound dressings on blood coagulation and platelet activation. *J Biomater Appl*. 1998;12(3):249–57.
140. Brown CD, Zitelli JA. Choice of wound dressings and ointments. *Otolaryngol Clin North Am*. 1995;28(5):1081–91.
141. Silverlon TM CA Antimicrobial Calcium Alginate Dressing. FDA Approval Letter. 510 (k) Number: K053590. 2006.
142. Bracey A, Shander A, Aronson S, Boucher BA, Calcaterra D, Chu MW, et al. The use of topical hemostatic agents in cardiothoracic surgery. *Ann Thorac Surg*. 2017;104(1):353–60.
143. Lapiere F, D'Houtaud S, Wager M. Hemostatic agents in neurosurgery. Explicative cases of controversial issues in neurosurgery: IntechOpen; 2012.
144. ETHICON OMNEX SURGICAL SEALANT®, Premarket Approval by FDA. PMA Number: P060029. 2010.
145. Hagberg RC, Safi HJ, Sabik J, Conte J, Block JE. Improved intraoperative management of anastomotic bleeding during aortic reconstruction: results of a randomized controlled trial. *Am Surg*. 2004;70(4):307–11.
146. Tan C, Utley M, Paschalides C, Pilling J, Robb JD, Harrison-Phipps KM, et al. A prospective randomized controlled study to assess the effectiveness of CoSeal® to seal air leaks in lung surgery. *Eur J Cardiothorac Surg*. 2011;40(2): 304–8.
147. Köhler C, Kyeyamwa S, Marnitz S, Tsunoda A, Vercelino F, Schneider A, et al. Prevention of lymphoceles using FloSeal and CoSeal after laparoscopic lymphadenectomy in patients with gynecologic malignancies. *J Minim Invasive Gynecol*. 2015;22(3):451–5.
148. Coseal Surgical Sealant®, Premarket Approval by FDA. PMA Number: P030039. 2018.
149. Duraseal Dural Sealant System®, Premarket Approval by FDA. PMA Number: P040034. 2018.
150. Nadler RB, Loeb S, Rubenstein RA, Vardi IY. Use of BioGlue in laparoscopic partial nephrectomy. *Urology*. 2006;68(2):416–8.
151. LeMaire SA, Schmittling ZC, Coselli JS, Ündar A, Deady BA, Clubb FJ Jr, et al. BioGlue surgical adhesive impairs aortic growth and causes anastomotic strictures. *Ann Thorac Surg*. 2002;73(5):1500–6.
152. BioGlue® Surgical Adhesive, FDA Approval Letter. H990007. 1999.
153. Makower J, Chang JY, Muni KP, Carlyle W, Levine H, Facticeau WM. Mucosal tissue dressing and method of use. Google Patents: US9381270B2. 2016.
154. Hanes RM, Hanes AL. Natural polymer based tissue adhesive with healing promoting properties. Google Patents: US20190038798A1. 2019.
155. Mannucci PM. Hemostatic drugs. *N Engl J Med*. 1998;339(4):245–53.
156. Savva A, Taylor MJ, Beatty CW. Management of cerebrospinal fluid leaks involving the temporal bone: report on 92 patients. *Laryngoscope*. 2003; 113(1):50–6.
157. Maxwell JA, Goldware SI. Use of tissue adhesive in the surgical treatment of cerebrospinal fluid leaks: experience with isobutyl 2-cyanoacrylate in 12 cases. *J Neurosurg*. 1973;39(3):332–6.
158. Spangler D, Rothenburger S, Nguyen K, Jampani H, Weiss S, Bhende S. In vitro antimicrobial activity of oxidized regenerated cellulose against antibiotic-resistant microorganisms. *Surg Infect (Larchmt)*. 2003;4(3):255–62.
159. Wiseman D, Kamp L, Saferstein L, Linsky C, Gottlick L, Diamond MP. Improving the efficacy of INTERCEED barrier in the presence of blood using thrombin, heparin or a blood insensitive barrier, modified INTERCEED (nTC7). *Prog Clin Biol Res*. 1993;381:205.
160. Wormald P, Sellars S. Bismuth subgallate: a safe means to a faster adeno—tonsillectomy. *J Laryngol Otol*. 1994;108(9):761–2.
161. Callanan V, Curran AJ, Smyth DA, Gormley PK. The influence of bismuth subgallate and adrenaline paste upon operating time and operative blood loss in tonsillectomy. *J Laryngol Otol*. 1995;109(3):206–8.
162. Hatton RC. Bismuth subgallate—epinephrine paste in adenotonsillectomies. *Ann Pharmacother*. 2000;34(4):522–5.
163. Sørensen W, Henrichsen J, Bonding P. Does bismuth subgallate have haemostatic effects in tonsillectomy? *Clin Otolaryngol Allied Sci*. 1999; 24(1):72–4.
164. Murray AD, Gibbs SR, Billings KR, Biavati MJ. Respiratory difficulty following bismuth subgallate aspiration. *Arch Otolaryngol Head Neck Surg*. 2000; 126(1):79–81.
165. Falbe-Hansen J, Jacobsen B, Lorenzen E. Local application of an antifibrinolytic in tonsillectomy a double-blind study. *J Laryngol Otol*. 1974; 88(6):565–8.
166. Robb P, Thorning G. Perioperative tranexamic acid in day-case paediatric tonsillectomy. *Ann R Coll Surg Engl*. 2014;96(2):127–9.
167. CYKLOKAPRON®tranexamic acid injection. FDA Approval Letter. 1999.
168. Carter G, Goss A. Tranexamic acid mouthwash—a prospective randomized study of a 2-day regimen vs 5-day regimen to prevent postoperative bleeding in anticoagulated patients requiring dental extractions. *Int J Oral Maxillofac Surg*. 2003;32(5):504–7.
169. Costin J, Ansell J, Lailicht B, Bakhru S, Steiner S. Reversal agents in development for the new oral anticoagulants. *Postgrad Med*. 2014; 126(7):19–24.
170. Sindet-Pedersen S, Stenbjerg S. Effect of local antifibrinolytic treatment with tranexamic acid in hemophiliacs undergoing oral surgery. *J Oral Maxillofac Surg*. 1986;44(9):703–7.
171. Upadhyay SP, Mallick PN, Jagia M, Singh RKA. Acute arterial thrombosis associated with inadvertent high dose of tranexamic acid. *Indian J Crit Care Med*. 2013;17(4):237.
172. Teker AM, Korkut AY, Kahya V, Gedikli O. Prospective, randomized, controlled clinical trial of Ankaferd blood stopper in patients with acute anterior epistaxis. *Eur Arch Otorhinolaryngol*. 2010;267(9):1377–81.

173. Kaya FS, Akova YA. The effect of autologous serum eye drop application on epithelization in the treatment of various ocular surface disorders and its safety/Cesitli nedenlere bagli goz yuzeyi sorunlarinda topikal otolog serum damla uygulamasinin epitelizasyona etkisi ve guvenilirliigi. *Turkish J Ophthalmol*. 2012;42(5):336–42.
174. Matsuo A, Yamazaki Y, Takase C, Aoyagi K, Uchinuma E. Osteogenic potential of cryopreserved human bone marrow-derived mesenchymal stem cells cultured with autologous serum. *J Craniofac Surg*. 2008;19(3):693–700.
175. Antisdell JL, West-Denning JL, Sindwani R. Effect of microporous polysaccharide hemospheres (MPH) on bleeding after endoscopic sinus surgery: randomized controlled study. *Otolaryngol Head Neck Surg*. 2009;141(3):353–7.
176. Antisdell JL, Janney CG, Long JP, Sindwani R. Hemostatic agent microporous polysaccharide hemospheres (MPH) does not affect healing or intact sinus mucosa. *Laryngoscope*. 2008;118(7):1265–9.
177. Zucker W, Mason R. Ultrastructural aspects of interactions of platelets with microcrystalline collagen. *Am J Pathol*. 1976;82(1):129.
178. Singh R, Shitiz K, Singh A. Chitin and chitosan: biopolymers for wound management. *Int Wound J*. 2017;14(6):1276–89.
179. Nakajima M, Kamei T, Tomimatu K, Manabe T. An intraperitoneal tumor mass caused by granulomas of microfibrillar collagen hemostat (Avitene). *Arch Pathol Lab Med*. 1995;119(12):1161–3.
180. Kulkarni MR. The use of platelet-rich fibrin as a hemostatic material in oral soft tissues. *Oral Maxillofac Surg*. 2019;23(1):119.
181. Raja VS, Naidu EM. Platelet-rich fibrin: evolution of a second-generation platelet concentrate. *Indian J Dent Res*. 2008;19(1):42.
182. Ishihara M. Photocrosslinkable chitosan hydrogel as a wound dressing and a biological adhesive. *Trends Glycosci Glycotechnol*. 2002;14(80):331–41.
183. Yusof NLBM, Lim LY, Khor E. Preparation and characterization of chitin beads as a wound dressing precursor. *J Biomed Mater Res*. 2001;54(1):59–68.
184. Jia Z, Xu W. Synthesis and antibacterial activities of quaternary ammonium salt of chitosan. *Carbohydr Res*. 2001;333(1):1–6.
185. Okamoto Y, Kawakami K, Miyatake K, Morimoto M, Shigemasa Y, Minami S. Analgesic effects of chitin and chitosan. *Carbohydr Polym*. 2002;49(3):249–52.
186. Shanmugapriya K, Kim H, Saravana PS, Chun B-S, Kang HW. Fabrication of multifunctional chitosan-based nanocomposite film with rapid healing and antibacterial effect for wound management. *Int J Biol Macromol*. 2018;118:1713–25.
187. Kean T, Thanou M. Biodegradation, biodistribution and toxicity of chitosan. *Adv Drug Deliv Rev*. 2010;62(1):3–11.
188. Hu Y, Zhang Z, Li Y, Ding X, Li D, Shen C, et al. Dual-crosslinked amorphous polysaccharide hydrogels based on chitosan/alginate for wound healing applications. *Macromol Rapid Commun*. 2018;39(20):1800069.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

